

# When Darwin's Special Difficulty Promotes Diversification in Insects

Frédéric Legendre, Fabien Condamine

### ▶ To cite this version:

Frédéric Legendre, Fabien Condamine. When Darwin's Special Difficulty Promotes Diversification in Insects. Systematic Biology, 2018, 67 (5), pp.873-887. 10.1093/sysbio/syy014. mnhn-02170200

## HAL Id: mnhn-02170200 https://mnhn.hal.science/mnhn-02170200v1

Submitted on 1 Jul2019

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1	Running head
2	EUSOCIALITY AND DIVERSIFICATION
3	
4	Title
5	When Darwin's Special Difficulty Promotes Diversification in Insects
6	
7	Authors' names
8	Frédéric Legendre <sup>1¶</sup> * & Fabien L. Condamine <sup>2¶</sup> *
9	
10	Authors' affiliations
11	<sup>1</sup> Muséum national d'Histoire naturelle, UMR 7205 Institut de Systématique, Evolution,
12	Biodiversité ISYEB – UMR 7205 – MNHN CNRS UPMC EPHE, Sorbonne Universités, 57
13	rue Cuvier, 75231 Paris Cedex 05, France
14	<sup>2</sup> CNRS, UMR 5554 Institut des Sciences de l'Evolution (Université de Montpellier), Place
15	Eugène Bataillon, 34095 Montpellier, France
16	
17	*Correspondence to be sent to: Muséum national d'Histoire naturelle, UMR 7205 Institut de
18	Systématique, Evolution, Biodiversité ISYEB – UMR 7205 – MNHN CNRS UPMC EPHE,
19	Sorbonne Universités, 57 rue Cuvier, 75231 Paris Cedex 05, France; E-mail:
20	frederic.legendre@mnhn.fr; CNRS, UMR 5554 Institut des Sciences de l'Evolution
21	(Université de Montpellier), Place Eugène Bataillon, Bât. 22 RDC, CC064, 34095
22	Montpellier, France; E-mail: fabien.condamine@gmail.com.
23	
24	<sup>¶</sup> Both authors contributed equally to this work.
25	

26 Abstract. – Eusociality, Darwin's special difficulty, has been widely investigated but remains 27 a topic of great debate in organismal biology. Eusocial species challenge existing theories, 28 and the impact of highly integrated societies on diversification dynamics is controversial with 29 opposing assertions and hypotheses in the literature. Here, using phylogenetic approaches in 30 termites - the first group that has evolved eusociality - we assessed the fundamental 31 prediction that eusocial lineages have higher diversification rates than non-eusocial clades. 32 We found multiple lines of evidence that eusociality provided higher diversification as 33 compared to non-eusociality. This is particularly exacerbated for eusocial species with 'true' 34 workers as compared to species with 'false' workers. Because most species with 'true' 35 workers have an entirely prokaryotic microbiota, the latter feature is also related to higher 36 diversification rates, but it should be investigated further, notably in relation to angiosperm 37 diversification. Overall, this study suggests that societies with 'true' workers are not only 38 more successful at ecological timescales but also over millions of years, which further implies 39 that both organism- and species-level traits act on species selection.

40

41 *Keywords.* –Aggregate trait, emergent trait, eusocial insects, macroevolution, species

42 selection, termites

Eusociality is the most integrated organizational level of animal sociality and is 44 45 defined by cooperative brood care, overlapping generations within a colony of adults, and 46 division of labour into reproductive and non-reproductive groups (Wilson and Hölldobler 47 2005). Although the origin of eusociality is still widely investigated and debated (e.g. Nowak et al. 2010; Nonacs 2011; Rousset and Lion 2011; Johnstone et al. 2012), this rare but 48 49 widespread behavioural characteristic has independently appeared in insects (at least 7-20 50 times depending on the definition of eusociality), crustaceans (at least thrice), and mammals 51 (at least twice) (Fig. 1; Bourke 2011). Within insects, eusocial species have colonies with 52 caste differences: queens and reproductive males take the roles of sole reproducers, while 53 soldiers defending the colony and workers foraging and maintaining resources contribute to 54 creating a living situation favourable for the colony (Wilson 1971). In his theory of natural 55 selection, Darwin (1859) famously portrayed the eusocial insects as a "special difficulty, 56 which at first appeared to [him] insuperable and actually fatal to [his] theory". Indeed, 57 eusociality presents an apparent paradox: if adaptive evolution is mediated by differential 58 reproduction of individuals on which natural selection acts, how can individuals incapable of 59 passing on their genes evolve and persist?

60

Figure 1. Origin and timeline of eusocial lineages for mammals, crustaceans and insects (bees, wasps, ants, and termites). Bars represent temporal ranges with solid lines based on the fossil record, whereas broken lines indicate possible eusociality supported by molecular dating studies (ants: Moreau et al. 2006; bees: Cardinal and Danforth 2011, wasps: Hines et al. 2007, termites: Ware et al. 2010; Legendre et al. 2015b) but not supported by fossil evidence.

68 The multilevel selection theory could settle this paradox. This theory has a long and 69 troubled history (Okasha 2006) but is increasingly accepted and supported, especially at the 70 species level (Jablonski 2008). Species selection results from differences in speciation and 71 extinction rates among species within a clade, and these differences are not merely related to 72 the fitness at the level of individuals (Jablonski 2008; Rabosky and McCune 2010). Data and 73 theory suggest that both organism-level and species-level traits (called aggregate and 74 emergent traits, respectively) act on species selection. Aggregate traits are characteristics of 75 individual organisms (e.g. body size), whereas emergent traits are species properties that are 76 not reducible to organismal traits such as eusociality. Extending this vocabulary, authors often 77 refer to the emergent fitness of species within clades when discussing species selection 78 (Jablonski 2008).

79 At the macroevolutionary timescale, the origin and evolution of eusociality are 80 increasingly well-understood due to recent phylogenetic analyses, along with estimates of 81 divergence times and diversification rates notably for arthropod groups (ants: Moreau et al. 82 2006; Pie and Feitosa 2016, bees: Cardinal and Danforth 2011, beetles: Farrell et al. 2001, 83 wasps: Hines et al. 2007, spiders: Agnarsson et al. 2006, termites: Ware et al. 2010). These 84 studies have investigated the number of origins of eusociality and the putative factors behind 85 the extraordinary diversification of eusocial groups (e.g. the Cretaceous radiation of 86 Angiosperms: Moreau et al. 2006; Cardinal and Danforth 2013). But another challenge 87 surrounds eusocial organism evolution: the impact of highly integrated societies on the 88 dynamics of diversification in these organisms (Davis et al. 2009; Ware et al. 2010). This 89 topic has received less attention probably due to a lack of methods to handle large 90 phylogenies that include both non-eusocial and eusocial lineages. Although the role of 91 eusociality has not been formally studied with state-of-the-art diversification methods, the 92 literature is filled with opposing assertions and hypotheses. In terms of ecological and

93 evolutionary success, modern eusocial insects can either be seen as species-rich groups such 94 as ants, bees, or termites (Chapman and Bourke 2001; Wilson 1990) or as a group with lower 95 species diversity than non-eusocial relatives such as ambrosia beetles, or poplar spiral gall 96 aphids (Wilson 1992). Although the success of eusocial groups could naively lead us to think 97 that eusociality was a trigger of diversification, Wilson (1992) argued that social insects 98 (contrary to social vertebrates) show lower diversification (because they are "less speciose") 99 than non-eusocial insects. Based on studies of population genetics and molecular evolution, 100 sociality reduces effective population size and possibly reduces levels of genetic variation, 101 which would in turn increase the risk of extinction of social organisms (Lanfear et al. 2014). 102 Hence, eusociality could indeed be seen as an impediment to diversification. The fossil record 103 shows that taxa went extinct within eusocial groups (ants: LaPolla et al. 2013, termites: 104 Krishna et al. 2010) but, so far, eusocial insect groups have been successful at the 105 macroevolutionary scale. For the most part, they have persisted since the origins of these 106 highly integrated behaviours, leading Wilson (1990) to state that "eusociality conveys 107 evolutionary long life to social insects". In reality, several emergent traits influence both 108 extinction and speciation rates, resulting in unpredictable trade-offs (Jablonski 2008). 109 The ensuing questions 'does eusociality confer higher diversification rates than non-110 eusocial species?' and 'what is the dynamic of diversification of social insects and what factor 111 could explain their success and potential shift in diversification rates?' are long-standing 112 research topics (Darwin 1859; Wilson 1992; Wilson and Hölldobler 2005). Biotic and abiotic 113 factors are important in diversification processes (Benton 2009) and range from 114 climatic/geologic events (e.g. Condamine et al. 2013), to competition (e.g. Liow et al. 2015; 115 Silvestro et al. 2015a), and key innovations (e.g. Rainford et al. 2014; Sánchez-García and 116 Matheny 2016). Eusociality can be classified as a key innovation, but has surprisingly been 117 understudied when one considers the ecological success that eusocial insect species have

experienced (Wilson 1971, 1992; Chapman and Bourke 2001). This ecological dominance is
mainly explained by one property of eusociality: the reproductive division of labour. In other
words, some individuals do not reproduce (i.e. they belong to sterile castes), and this has been
qualified as the most important feature of eusociality (Wilson 1990).

122 Within insects, termites (Dictyoptera, order Blattodea, infraorder Isoptera) contain 123 only eusocial species, and it is thought that they were the first modern eusocial animals to 124 evolve, sometime in the Late Jurassic (ca. 155 Ma) (Legendre et al. 2015b; Engel et al. 2016). 125 A termite colony is usually differentiated into reproductive, worker and soldier castes (Fig. 126 2a). The latter two castes, which are sterile and perform highly specialized tasks, derive from 127 different stages depending on the species (Noirot and Pasteels 1987). Whereas soldiers likely 128 have a single origin, the evolution of workers is more controversial (Watson and Sewell 1981; 129 Noirot and Pasteels 1987; Thompson et al. 2000; Grandcolas and D'Haese 2002; Inward 130 2007, Legendre et al. 2008, 2013; Roisin and Korb 2011). Importantly, two types of workers 131 were distinguished early: Some species have a 'true' worker caste, while others have a 'false' 132 worker caste (also called pseudergates; Fig. 2a). 'True' workers differ from pseudergates 133 notably because they diverge early and irreversibly from the imaginal line (Noirot and 134 Pasteels 1987), while pseudergates do not. Societies with 'true' workers have been qualified 135 as more 'socially' highly integrated, which might have consequences for their ecological and 136 evolutionary success given that >80% of all termites have 'true' workers (Korb 2009; Brune 137 2014).

138

Figure 2. Eusociality in termites: life cycle (a) and (b) evolutionary origin associated to
putative key innovations. The life cycle differs for species with pseudergates and species with
'true workers' (highlighted by the dashed box). The phylogeny of termites (Isoptera)
illustrates that the origin of eusociality is nested within the radiation of cockroaches (termites)

and cockroaches together forming the Blattodea). The phylogeny also highlights important
events in the evolution of the digestive symbiosis: the presence of cellulolytic flagellates,
acquired by a common ancestor of termites and Cryptocercidae, is associated to their woodfeeding lifestyle; the loss of flagellates in Termitidae gave rise to an enormous dietary
diversification (chronogram adapted from Legendre et al. 2015b). Dashed lines for the
common ancestor of cockroaches denote that the group is older than 201 Ma. The asterisk (\*)
specifies that some families or subfamilies are paraphyletic according to the latest studies.

151 The study of termite digestive systems might also bring important insights on the 152 ecological and evolutionary successes of these insects (Fig. 2b). Although digestive strategies 153 and the gut microbiota of termites remain to be fully deciphered, important progress has been 154 made and two groups can be distinguished by their primary cellulolytic partners and their 155 hindgut compartmentation (Brune 2014). The first group comprises termite species that 156 harbour cellulolytic flagellates in their gut; they group all termite species except those 157 belonging to Termitidae. In the second group (i.e. Termitidae), these flagellates are absent; 158 instead Termitidae have an entirely prokaryotic microbiota. They also show a higher 159 compartmentation of the hindgut (except for Macrotermitinae) and a large dietary 160 diversification (Brune and Dietrich 2015). Because most termite species belong to the family 161 Termitidae (Beccaloni and Eggleton 2013), their peculiar digestive system might have been a 162 trigger of diversification. A parallel can be made with cockroach species; even though the gut 163 microbiota of most cockroach species has not been investigated, flagellates have been found 164 in some wood-eating species, including, but not restricted to (e.g. Pellens et al. 2002), 165 *Cryptocercus*, the alleged sister group of termites (Cleveland et al. 1934; Klass et al. 2008). 166 Because of characteristics shared with 'lower' termites (Klass et al. 2008), it is then generally 167 assumed that Cryptocercus belongs to the group with cellulolytic flagellates in their gut

(Brune and Dietrich 2015), contrary to other supposedly or asserted wood-eating species (but
see Pellens et al. 2007 and the section *Trait-dependent diversification* for more details and
alternative codings).

171 Our knowledge on the role of eusociality in the dynamic of diversification is limited. 172 Only one previous study has investigated the diversification of termites (Davis et al. 2009), 173 suggesting increases in diversification early in termite evolution. However, this study was 174 based on a family-level supertree, and methods to estimate diversification were still in their 175 infancy. Besides, this study did not differentiate the potential role of 'true' workers or that of 176 gut microbiota composition in termite diversification. To better understand the triggers of 177 diversification in Dictyoptera (mantises, cockroaches and termites), we analyze the most 178 comprehensive time-calibrated phylogeny of the group. We investigate the diversification 179 dynamic of termites compared to their dictyopteran counterparts using a battery of birth-death 180 methods to corroborate and strengthen these results. More specifically, we tested the 181 competing hypotheses in which eusociality is associated with lower (Wilson 1992) or higher 182 (Davis et al. 2009) net diversification rate compared to non-eusociality. We also tested 183 whether the presence of 'true' workers in termite societies can be considered a key innovation 184 relative to societies with pseudergates, by assessing the putative role on diversification of 185 these castes on termite evolution. Similarly, we tested whether an entirely prokaryotic gut 186 microbiota could also be a key innovation for termite diversification.

187

#### 188 MATERIALS AND METHODS

189 Analytical Pipeline

We used a recently published time-calibrated phylogeny of Dictyoptera including 762
species representatives of living dictyopteran diversity (*ca.* 10,000 known species, Legendre
et al. 2015b). This phylogeny resulted from a molecular dataset comprising four

193 mitochondrial and two nuclear markers. Divergence times were estimated using a relaxed-

194 clock approach coupled with 17 fossils, which were used as minimum age constraints on

different nodes and a maximum age for the tree root (Legendre et al. 2015b).

Three putatively key innovations were investigated: eusociality, societies with 'true'
workers and entirely prokaryotic gut microbiota. To test the role of these innovations, we used
three approaches: *(i)* the maximum likelihood (ML) approach of time-dependent

diversification (Morlon et al. 2011), implemented in the R-package *RPANDA* v.1.2 (Morlon et

al. 2016), (ii) the Bayesian analysis of macroevolutionary mixture (BAMM v.2.5, Rabosky et

al. 2013), implemented in the R-package *BAMMtools* v.2.1.4 (Rabosky et al. 2014), and *(iii)* 

the ML approach of trait-dependent diversification (Madisson et al. 2007; FitzJohn et al.

203 2009), implemented in the R-package *diversitree* v.0.9-8 (FitzJohn 2012).

204 Each method is designed to estimate speciation and extinction rates, and the three 205 methods are used to cross-test hypotheses and corroborate results. Nonetheless, it is worth 206 mentioning that each method differs at several points in the way speciation and extinction 207 rates are estimated. For instance, trait-dependent birth-death models (diversitree) estimate 208 constant speciation and extinction rates, while time-dependent birth-death models (BAMM 209 and RPANDA) estimate the speciation and extinction rates and their variation through time. 210 Therefore we expect some differences in the values of the estimated diversification rates that 211 are inherent to each approach. Our diversification analyses should be seen as complementary 212 for the inferred diversification trend rather than corroborative on the values and magnitude of 213 the speciation and extinction rates.

214

215 Across-Clade and Time-Variation Diversification

The ML approach of Morlon et al. (2011) is a birth-death method that extendsprevious birth-death methods such that speciation and/or extinction rates may change

continuously through time, and subclades may have different speciation and extinction rates.
This method has the advantage of not assuming constant extinction rate over time (unlike
BAMM, Rabosky et al. 2013), and allows clades to have declining diversity because
extinction can exceed speciation, meaning that diversification rates can be negative (Morlon
et al. 2011).

We designed six nested diversification models to test with this approach: *(i)* a Yule model, where speciation is constant and extinction is null; *(ii)* a constant birth-death model, where speciation and extinction rates are constant; *(iii)* a variable speciation rate model without extinction; *(iv)* a variable speciation rate model with constant extinction; *(v)* a rateconstant speciation and variable extinction rate model; and *(vi)* a model in which both speciation and extinction rates vary. Models were compared by computing the ML score of each model and the resulting Akaike information criterion corrected by sample size (AICc).

230 First, the Dictyoptera tree was analyzed as a whole using this approach and taking into 231 account missing species (f=0.08). To test the hypotheses that eusociality, societies with 'true' 232 workers and gut microbiota composition act on diversification rates, we compared the 233 likelihoods of models that allow for different patterns of rate variation in different clades. 234 More specifically, we first allowed for a rate shift along the branch corresponding to the 235 apparition of eusociality (the termite crown node), and we sequentially tested for a rate shift 236 between the crown node of termites and the node corresponding to the family Termitidae 237 (prokaryotic microbiota). Therefore, we partitioned the Dictyoptera phylogeny into seven 238 evolutionary scenarios: from a backbone without the termites, and the termites as a subtree; to 239 a backbone without the Termitidae, and the Termitidae as a subtree (with all five possibilities 240 in between; for a schematic illustration see Fig. S1 in the Supplementary Material that 241 accompanies this article available on Dryad at http://dx.doi.org/10.5061/dryad.2tg34). After 242 creating these backbones and corresponding subtrees, and accounting for the missing species

243 in each of them, we fitted the same diversification models as detailed above but independently 244 on each subtree and its corresponding backbone tree (Fig. S1 on Dryad). The global log-245 likelihood was calculated as the sum of both subclade and backbone log-likelihoods as 246 determined by the corresponding best-fitting models. This approach allowed us to compare 247 the global log-likelihood (under the assumption of a single speciation rate and extinction rate) 248 to the log-likelihood of one shift at the termite crown (eusociality), and to the log-likelihood 249 of one shift at the crown of Termitidae (prokaryotic microbiota). We directly compare the 250 seven scenarios using AICc.

251 We also used BAMM to estimate speciation and extinction rates through time and 252 among/within clades (Rabosky et al. 2013). BAMM was constructed to study complex evolutionary processes on phylogenetic trees, potentially shaped by a heterogeneous mixture 253 254 of distinct evolutionary dynamics of speciation and extinction across clades. BAMM can 255 automatically detect rate shifts and sample distinct evolutionary dynamics that explain the diversification dynamics of a clade without a priori hypotheses on how many and where these 256 257 shifts might occur. Evolutionary dynamics can involve time-variable diversification rates; in 258 BAMM, speciation is allowed to vary exponentially through time while extinction is 259 maintained constant: subclades in a tree may diversify faster (or slower) than others. This 260 Bayesian approach can be useful in detecting shifts of diversification potentially associated 261 with key innovations (Rabosky 2014).

We ran BAMM by setting four Markov chain Monte Carlo running (MCMC) for 20 million generations and sampled every 2,000 generations. A compound Poisson process is implemented for the prior probability of a rate shift along any branch. We used a gradient of prior values ranging from 0.1 to 50 to test the sensitivity to the prior, because it has been shown that BAMM can be affected by the prior (Moore et al. 2016 but see Rabosky et al. 2017). We accounted for non-random incomplete taxon sampling using the implemented

analytical correction; we set a sampling fraction for mantises (f=0.118), cockroaches

(f=0.042), and termites (f=0.094). We performed independent runs (with a 15% burn-in) using different seeds to assess the convergence of the runs with effective sample size. We processed the output data using the *BAMMtools* (Rabosky et al. 2014) by estimating (*i*) the mean global rates of diversification through time, (*ii*) the configuration of the diversification rate shifts evaluating alternative diversification models as compared by posterior probabilities, and (*iii*) the clade-specific rates through time when a distinct macroevolutionary regime is identified.

275

#### 276 Trait-Dependent Diversification

277 We used trait-dependent diversification models to simultaneously model trait evolution and its impact on diversification (Maddison et al. 2007). In those models, species 278 279 are characterized by an evolving trait, and their diversification follows a birth-death process in 280 which speciation and extinction rates may depend on the trait state. We created three datasets 281 of traits. First, we categorized all dictyopteran species as being non-eusocial or eusocial. This 282 two-trait scheme distinguishes the termites (all species are eusocial) from the rest of 283 dictyopterans (all non-eusocial). Second, we made a three-trait scheme by keeping the non-284 eusocial category, and dividing the eusocial trait into two: eusocial with pseudergates ('false' 285 workers) and eusocial with 'true' workers. Third, we made another three-trait scheme to test 286 the role of the primary cellulolytic partners in diversification. We followed Brune and 287 Dietrich (2015) to distinguish Termitidae (entirely prokaryotic microbiota), Cryptocercus + 288 other termites (cellulolytic flagellates), and the remaining Dictyoptera (no specialized 289 microbiota for lignocellulose digestion). All these coding schemes are imperfect because, like 290 any broad categorization process, they might oversimplify the reality found in nature 291 (Legendre et al. 2015a; Goutte et al. 2016). The category 'remaining Dictyoptera' in the latter 292 coding scheme, for instance, is imperfect because it is delineated by default and because of a

293 lack of information for some cockroach species. However, this three-state coding reflects our 294 current understanding of the primary cellulolytic partners in Dictyoptera as reviewed in Brune 295 and Dietrich (2015). Anyway, to take into account our lack of knowledge on cockroach 296 microbiota and the existence of species that, arguably, could not fit well in these three 297 categories – while limiting the number of categories to save statistical power, we created two 298 supplementary trait datasets. In the first alternative coding, the wood-feeding non-eusocial 299 lineages were coded with cellulolytic flagellates (same as 'lower' termites) even though the 300 existence and role of cellulolytic flagellates have not always been investigated and proved. 301 These lineages represent several genera of the subfamily Panesthiinae, and the genera 302 Cyrtotria, Lauraesilpha, Paramuzoa, Parasphaeria, and Colapteroblatta (Grandcolas 1993; 303 Grandcolas et al. 2002; Pellens et al. 2002). In the second alternative coding, the 304 Macrotermitinae were not coded with the Termitidae (i.e. initial coding supported by their 305 entirely prokaryotic gut microbiota) but with the other termites (coding supported by their 306 relatively simple hindgut structure).

307 We applied the Binary State Speciation and Extinction model (BiSSE, Maddison et al. 308 2007) with constant diversification rate for the two-trait dataset. We then performed the 309 Multi-State Speciation and Extinction model (MuSSE, FitzJohn et al. 2009) on the three-trait 310 datasets. Both BiSSE and MuSSE models account for incomplete taxon sampling, which is 311 informed as a sampling fraction of species at present having a given trait (FitzJohn et al. 312 2009). Simulation studies have shown that a large, well-sampled tree is required by these 313 methods, whereas trees containing fewer than 300 species may lack sufficient phylogenetic 314 signal to produce enough statistical power (Davis et al. 2013).

315 The BiSSE model has six distinct parameters: two speciation rates without character 316 change (i.e. *in situ* speciation) associated with non-eusocial species ( $\lambda_N$ ) and eusocial species 317 ( $\lambda_E$ ), two extinction rates associated with non-eusocial ( $\mu_N$ ) and eusocial species ( $\mu_E$ ), and two

transition rates (i.e. anagenetic change) with one from non-eusocial to eusocial ( $q_{N-E}$ ), and from eusocial to non-eusocial ( $q_{E-N}$ ). Using the same rationale, but applied to three traits, the MuSSE model has 12 distinct parameters (three for speciation, three for extinction, and six for transitions).

Here we used 100 dictyopteran phylogenies (sampled from the Bayesian analysis in Legendre et al. 2015b) and combined them with our three datasets of traits. Analyses were performed using the R-package *diversitree* (FitzJohn 2012) using the functions *make.bisse* and *make.musse* to construct the likelihood functions for each model based on the data, and the functions *constrain* and *find.mle* to apply different diversification scenarios.

For each model, we computed the AICc based on the log-likelihood and the number ofparameters. We checked support for the selected model against all models using the

329 difference between AICc ( $\Delta$ AIC) and the Akaike weight (AIC $\omega$ ). The scenario supported with

the lowest AICc was considered the best when  $\Delta AIC > 2$  against other models and with

 $AIC\omega > 0.5$  (otherwise the model with less parameter was instead considered the best). Finally,

332 we used the consensus tree and MCMC simulations with the best model to examine the

333 confidence interval of the parameter estimates. We used an exponential prior and started the

chain with the parameters obtained by maximum likelihood (FitzJohn 2012). We ran 20,000

335 MCMC steps and applied a 10% burn-in. Net diversification rates were then computed.

336 SSE models have recently been criticized due to type I error (Rabosky and Goldberg 337 2015). To test whether our diversification results were biased, we estimated the difference of 338 fit ( $\Delta$ AIC) between the best model and a null model (i.e. with no state dependence) and 339 compared this with the difference between the same models as estimated from 100 simulated

341

340

342 RESULTS

trait datasets for each SSE model.

#### 343 Diversification Across-Clade and Time

344 The BAMM analyses supported a model with six evolutionary regimes (i.e. five rate 345 shifts; Fig. S2 and Table S1 available on Dryad). The post burn-in posterior distribution 346 indicated that five shifts occur with a posterior probability (PP) of 0.54, and PP=0.33 and 0.10 347 for four and six shifts, respectively (less or more shifts occur with PP<0.05; Table S1 348 available on Dryad). The 95% credible set of shift configurations and marginal shift 349 probabilities strongly favoured a model that includes shifts along branches within termites 350 (Figs S3-S4 available on Drvad). The shift configurations within this credible set contained 351 also shifts at internal nodes in mantises and at a more derived position within cockroaches 352 (Blaberidae: Panesthiinae). The different shift configurations in the credible set allowed 353 quantifying uncertainty in placement of a termite shift (Fig. S4 available on Dryad). The shift 354 configurations sampled at the highest frequency always contained a shift within termites 355 (cumulative posterior probability of 0.62). The best configuration shift retained five core 356 shifts, one located within the termites (crown Termitidae), two within Mantodea and two 357 others within cockroaches (Fig. S5 available on Dryad). We repeated the analyses by 358 changing the Poisson process for the prior probability of a rate shift along any branch: these 359 analyses supported a similar diversification pattern, although we found an additional clade-360 specific macroevolutionary regime when a higher prior probability of rate shift was used (Fig. 361 S6 and Table S1 available on Dryad).

We found evidence for low and constant-rate diversification through time for Dictyoptera punctuated by few shifts. When diversification changed, speciation rates increased for the termites in particular at the base of Termitidae (**Fig. 3**). Although other increases of speciation were found, the most elevated speciation rate (and also net diversification rate) occurred within termites. Interestingly, when termite macroevolutionary regime was studied in isolation, the net diversification rate of the clade increased towards the

present (Fig. 3). On the contrary, the other macroevolutionary regimes showed a morecommon pattern of diversification slowdown over time.

370

371 Figure 3. Diversification and eusociality in Dictyoptera. (a) Time-calibrated phylogeny of 372 Dictyoptera (adapted from Legendre et al. 2015b) showing the origin of eusociality (all 373 termites), the origin of an entirely prokaryotic microbiota (Termitidae), and the shifts in 374 diversification rate identified with BAMM. (b) Results of BiSSE show that lineages with 375 eusocial species have higher net diversification rates than non-eusocial species. (c) Results of 376 MuSSE show that lineages with 'true' workers have higher net diversification rates than other 377 dictyopteran lineages, including termite lineages with pseudergates. (d) Results of MuSSE 378 show that lineages with an entirely prokaryotic microbiota (and a complex hindgut 379 compartmentation) have a higher diversification rate than other dictyopteran lineages. For 380 BiSSE and MuSSE plots, Bayesian posterior distributions represent the 95% credibility 381 interval of each estimated parameter. (e) BAMM analyses suggest a strong increase in net 382 diversification rate of termites towards the present, especially in termites with 'true' workers 383 (mostly Termitidae and Rhinotermitidae). The shaded areas represent the 95% credibility 384 interval of each estimated parameter.

385

#### 386 Termite Key Innovations and Diversification

387 The *RPANDA* analyses refined the location of the rate shift within termites, and 388 allowed testing the effect of three termite key innovations: eusociality, societies with 'true' 389 workers, and entirely prokaryotic gut microbiota, which have evolved sequentially from the 390 origin of termites to the origin of Termitidae. Applying a series of time-dependent birth-death 391 models (including a rate shift at the crown node of a termite subclade), we compared the fit of 392 alternative scenarios to locate the best change of diversification rates. We first found that any

393 scenario including a rate shift within termites is better than a scenario having no shift at all 394 (**Table 1**). Comparing all diversification scenarios including a shift in the termite tree, we 395 found that the best-fitting scenario includes a shift along a branch supporting termites that 396 only contain species with 'true' workers in their societies (*scenario 6*, **Table 1**). There is 397 strong support for this scenario (lowest  $\Delta AIC=12.7$ ) as compared to competing hypotheses of 398 other locations in the termite tree such as the evolution of eusociality (*scenario 1*), or the 399 change in gut microbiota composition (*scenario 7*).

400 BAMM identified an important rate shift within the mantises (166.8 Ma, Fig. 3), 401 which corresponds to an increase of diversification. Using RPANDA, we tested whether a 402 model including this shift performs better than the model with the best-fitting shift in termites. 403 We reproduced the approach explained above but we isolated the mantis clade as a subtree, 404 from the rest of the Dictyoptera (this time including termites). We analyzed all the same 405 models and found that the shift within the Mantodea does not improve the likelihood as 406 compared with the model including the shift within termites (AICc for the shift in 407 mantises=7640.83; resulting in a  $\triangle AIC=327$  between the best scenario including a shift within 408 termites and this scenario, Table 1).

409 RPANDA analyses endorsed BAMM results: The best-fit scenario indicated a 410 speciation rate increasing through time for both the termite tree and the backbone. 411 Nonetheless, we estimated an elevated speciation rate of 0.193 lineage/Myr for termites, 412 strikingly contrasting with the speciation rate of the remaining dictyopteran lineages (0.058 413 lineage/Myr). Moreover, we found the termite clade diversified with low or zero extinction 414 rates, again contrasting with the extinction rate for the rest of the tree (0.023 lineage/Myr). 415 Consequently, the net diversification rate of the termite tree is 5.5 times higher than the 416 remaining part of the dictyopteran tree (scenario 7, Table 1).

417 We corroborated the BAMM and RPANDA analyses with BiSSE and MuSSE applied 418 to a two-trait dataset and two three-trait datasets, respectively. We found that the best-fitting 419 BiSSE model was the one with different speciation and extinction, but with equal transition 420 rates. Simpler or more complex models were not supported ( $\Delta AIC=4.13$  with the second best-421 fit BiSSE model, which has one extra parameter, Table 2a, Table S2 available on Dryad). In 422 the best-fit model, both speciation and extinction rates were higher for eusocial species 423 (Table 2a), and the net diversification rate was two-fold higher for eusocial lineages (Fig. 3). 424 Bayesian MCMC analysis showed that speciation, extinction, and net diversification rates 425 were significantly different between the two traits (Fig. S7 available on Dryad). 426 We further investigated whether difference of diversification rates occurred within the 427 termites by splitting the eusociality trait into two (societies with pseudergates or societies with 428 'true' workers). We found that the best-fitting MuSSE model supported the most complex 429 model, in which all parameters are free. Simpler models were not supported ( $\Delta AICc=15.8$ 430 with the second best-fit MuSSE model, which has one less parameter, Table 2b, Table S2 431 available on Dryad). In the best-fit model, speciation and extinction rates were higher for 432 eusocial lineages with 'true' workers. Speciation and extinction rates were both low for the 433 two other traits (non-eusocial, and eusocial with pseudergates). The net diversification rate 434 was higher for eusocial lineages with 'true' workers (Fig. 3). The trait-dependent rates of 435 speciation, extinction, and net diversification for the eusocial species with 'true' workers were 436 significantly different from the rates of the two other traits, and the two other traits were not 437 significantly different from each other (Fig. S8 available on Dryad). 438 As for the primary cellulolytic partners, we found that the net diversification rate was 439 higher for the Termitidae, the lineage with an entirely prokaryotic gut microbiota (Fig. 3;

440 **Table 2c**; Fig. S9 and Table S2 available on Dryad). The two other traits were not

significantly different in terms of diversification even though the lineages with cellulolytic

flagellates showed higher speciation and extinction rates than mantises and cockroaches(except *Cryptocercus* that harbours flagellates).

Randomization analyses of the three datasets of traits showed that all the SSE-based
results were robust to type-I error (Fig. S10 available on Dryad) indicating that our inferences
are not biased. We performed additional MuSSE analyses by re-coding either the woodfeeding non-eusocial lineages or Macrotermitinae with *Cryptocercus* spp. and all the termites
but the other Termitidae, and we found that the results were not sensitive to this effect:
eusocial lineages with a prokaryotic microbiota or with a complex hindgut compartmentation
diversified significantly faster than their counterparts (Figs. S11-S12 available on Dryad).

451

452 DISCUSSION

453 The origin of eusociality has been widely investigated at the organismal level. For 454 instance, Hall and Goodisman (2012) explored the differences in rate of substitution for 455 queen- and worker-selected loci, whereas Rehan and Toth (2015) reviewed several 456 hypotheses about molecular evolution of eusociality and tried to integrate them in a synthetic 457 framework. Above the species level, the origin and evolution of eusociality has been also 458 addressed with molecular dated phylogenies of eusocial groups (Farrell et al. 2001; 459 Agnarsson et al. 2006; Moreau et al. 2006; Hines et al. 2007; Cardinal and Danforth 2011, 460 2013; Pie and Feitosa 2016). These studies have shown that eusociality is a labile trait that has 461 appeared independently in many clades. However, the consequences of eusociality as a driver 462 of species diversification have received less attention (Davis et al. 2009; Ware et al. 2010). 463 What role does eusociality play at this macroevolutionary scale? In social insects, only two studies mentioned this aspect and suggested contradictory hypotheses. Edward O. Wilson 464 465 (1992) proposed that social insects would have a lower diversification rate than non-eusocial 466 insects, while Davis et al. (2009) suggested that termites would have a higher diversification

467 rate than their close relatives; the study of Ware et al. (2010) did not compare termite 468 diversification with the non-eusocial clades. These opposite predictions are in fact not so 469 surprising because, for numerous aggregate and emergent traits, both low speciation and 470 extinction rates are predicted (Jablonski 2008). These traits would then lead either to increase 471 or decrease in diversification depending on the intensity of speciation and extinction. 472 Speciation rate supposedly increases with molecular rate (Webster et al. 2003; Lanfear 473 et al. 2010; but see Rabosky and Matute 2013), which in turn depends on several features 474 including population size, population abundance, and genetic population. Harvey et al. (2017) 475 showed, for instance, a link between population differentiation within species and speciation 476 rates inferred from phylogenies in New World birds. All the aforementioned features are 477 emergent traits connected to eusociality. In termites, the archetype society mode involves 478 large and abundant populations headed by pairs of reproductives, resulting in colonies with 479 strong genetic structures and genetically isolated by distance (Goodisman and Crozier 2002; 480 Thompson et al. 2007; Dupont et al. 2009; Eggleton 2011; Vargo and Hussedener 2011). 481 According to classical predictions, high abundance would imply low extinction and low 482 speciation rates, while highly structured populations would imply both high extinction and 483 speciation rates. As for large population size, predictions suggest low extinction rates and are 484 contradictory about speciation rates (Jablonski 2008). For Dictyoptera, we found here higher 485 extinction and speciation rates for eusocial lineages when compared to non-eusocial lineages 486 (using three different approaches). Because speciation is much higher than extinction, the net 487 diversification rate is higher for eusocial species than for non-eusocial species, resulting in a 488 strong positive link between eusociality and net diversification.

These results suggest that a high level of genetic structure could be the main factor
acting on Dictyoptera diversification. However, the classical vision of termites is outdated.
Population structure in termites probably shows a large spectrum of variation that should be

492 investigated further (Vargo and Husseneder 2011) and compared to cockroach population 493 structures that should also vary between solitary, gregarious and subsocial species (Bell et al. 494 2007), or flying and wingless species. Population size – and size-dependent properties – is 495 also poorly known (Korb 2009) but range from a few hundreds to millions of individuals. 496 Even within the family Termitidae comprising only species with 'true' workers, there is a 497 large variance with nest population of a few thousands to a few millions of individuals 498 (Lepage and Darlington 2000). But, even though the underlying factors remain to be 499 investigated further, eusociality seems to act as a key innovation favouring diversification in 500 Dictyoptera.

501 Key innovations are alluded to whenever an evolutionary novelty is thought to have 502 favoured the diversification of a clade. Three main categories of key innovations can be 503 distinguished (Heard and Hauser 1995): (i) innovations opening new adaptive zones sensu 504 Simpson (1944); *(ii)* innovations increasing fitness and allowing to outcompete other species; 505 and *(iii)* innovations increasing the propensity for reproductive or ecological specialization. 506 These three categories are not necessarily exclusive. For termites, eusociality unlikely opened 507 new adaptive zones (definition 1). Termites act mainly as decomposers, a niche already 508 existing and occupied before the emergence of this clade (Raymond et al. 2000). However, a 509 highly integrated society, especially with 'true' workers, might have allowed termites to 510 outcompete other species and increased their propensity for reproductive and ecological 511 specialization (definitions 2 and 3). This scenario is classically put forward at the ecological 512 timescales, wherein the higher dominance and the large ecological success of termites are 513 known, and could be here translated at the macroevolutionary timescale. Interestingly, all the 514 diversification analyses (three different types of models) show that both speciation and 515 extinction rates were higher for eusocial lineages with 'true' workers resulting in a higher net 516 diversification rate for these lineages. Accordingly, our results suggest that societies with

517 'true' workers are not only more successful at ecological timescales but also over millions of
518 years, which further implies that both organism- and species-level traits (aggregate and
519 emergent traits, respectively) act on species selection.

520 Other factors are also probably at play in Dictyoptera diversification and they should 521 be examined in future research (Jablonski 2008; Benton 2009). The diversification of 522 angiosperms in the Cretaceous is often proposed to have played an important role in insect 523 diversification, especially social insects (Moreau et al. 2006; Cardinal and Danforth 2013). 524 Interestingly, molecular dating phylogenies of termites concur to show that termites appeared 525 in the very Late Jurassic – Early Cretaceous (Ware et al. 2010; Bourguignon et al. 2015; 526 Legendre et al. 2015b). Although there is a debate on the origin of angiosperms, from the 527 Early Cretaceous to Triassic (e.g. Beaulieu et al. 2015; Magallón et al. 2015; Silvestro et al. 528 2015b; Foster et al. 2017), the main period of diversification always occurred in the 529 Cretaceous and coincides well with the rise of many extant insect clades. It is nonetheless 530 questioned whether the angiosperm radiation has boosted insect diversification (Rainford and 531 Mayhew 2015; Condamine et al. 2016). The increase in termite diversification in the 532 Cretaceous can be associated with the rise to dominance of angiosperms; a similar scenario is 533 proposed for another important eusocial insect clade, the ants (Moreau et al. 2006). Other 534 dramatic landscape modifications that happened in the last 300 million years might have 535 promoted shifts in diversification in termites. For instance, global climate changes or the 536 break-up of ancient continents (Bourguignon et al. 2015) could have induced different 537 dynamics of diversification between non-eusocial and eusocial lineages. Future studies could 538 investigate the impact of such important environmental changes using both phylogenies and 539 fossil data. Also, to better understand these intricate mechanisms, approaches integrating 540 various factors such as caste and colony size but also foraging types, dispersal abilities, diets 541 and digestive capacity (Abe 1987; Jeanson and Deneubourg 2009; Brune and Dietrich 2015),

542 should be used to sort out the effect of these factors. Regarding diet specialization, which is 543 strongly related to gut anatomy and microbiota composition (Bignell and Eggleton 1995; 544 Köhler et al. 2012; Mikaelyan et al. 2015), the concomitance of termite and angiosperm 545 diversifications brings credit to its importance in termite diversification because changes in 546 intestinal anatomy and microbiota complexity would have opened new niches to diversify in. 547 Rabosky (2009) underlined the possible change in species "carrying capacity" for 548 some clades in a given area. Instead of playing on diversification rates *per se*, may eusociality 549 impact ecological limits in a given environment, which would in turn increase termite 550 diversification? This remains a possibility that could be linked to our side-result that net 551 diversification rates within termites increase towards the present, which is unusual, but the 552 underlying mechanisms are still to be deciphered. The apparent increased diversification rate 553 in termites at the present can be caused by a phenomenon, called the pull of the recent, which 554 is due to a more complete sampling of recent and still extant species than in the past 555 (Jablonsky 2008). This is slightly different from the pull of the present that is due to constant 556 birth-death models showing an upward turn in species number towards the present (Nee 557 2006). Most of the phylogenies are better explained by a slowdown of speciation rates 558 towards the present (e.g. Phillimore and Price 2008; Morlon et al. 2010). An increase in 559 speciation through time is rarely inferred, thus the increase in termite speciation goes against 560 the classical pattern. We do not think the increase in speciation can be attributed to the pull of 561 the present/recent because the increase started tens of million years ago, while the effect of 562 the pull of the present/recent is usually considered to be important in the last million years 563 (Nee 2006). The species sampling is relatively homogeneous across the phylogenetic tree, 564 except for the Panesthiinae which are more sampled than the rest, but more importantly the 565 termite subtree is not better sampled than the other groups (Legendre et al. 2015b). Therefore, 566 we think the impact of the pull of the present/recent is limited here.

567	Finally, even though there were probably multiple origins of the 'true' worker caste in
568	termites (Legendre et al. 2008, 2013), Dictyoptera offers a single replicate to investigate the
569	role of eusociality in clade diversification. We expect advances in our understanding of the
570	role of eusociality at the species selection level, when similar studies would be conducted
571	with other eusocial organisms, especially in the insect order Hymenoptera (Peters et al. 2017)
572	where ants, bees and wasps constitute conspicuous elements of ecosystems and have evolved
573	eusociality independently.
574	
575	SUPPLEMENTARY MATERIAL
576	Supplementary material can be found in the Dryad Digital Repository at http://datadryad.org,
577	http://dx.doi.org/10.5061/dryad.2tg34.
578	
579	ACKNOWLEDGMENTS
580	We thank the associate editor Brian Wiegmann and two anonymous referees for the excellent
581	comments that improved the study. We also thank Giovanni Fagua, Gaël Kergoat, Hélène
582	Morlon, André Nel and Felix Sperling for proofreading and commenting on this manuscript.
583	
584	Funding
585	Financial support was provided by a Marie Curie International Outgoing Fellowship (project
586	627684 BIOMME) to FLC.
587	
588	References
589	Abe T. 1987. Evolution of life types in termites. In: Evolution and Coadaptation in Biotic
590	Communities (eds Kawano, S., Connell, J.H., Hidaka, T.). Tokyo: University of
591	Tokyo Press, pp. 125–148.

- Agnarsson I., Avilés L., Coddington J.A., Maddison W.P. 2006. Sociality in theridiid spiders:
  repeated origins of an evolutionary dead end. Evolution 60:2342-2351.
- 594 Beaulieu J.M., O'Meara B.C., Crane P., Donoghue M.J. 2015. Heterogeneous rates of
- molecular evolution and diversification could explain the Triassic age estimate forangiosperms. Syst. Biol. 64:869-878.
- 597 Beccaloni G.W., Eggleton, P. 2013. Order Blattodea. Zootaxa 3703:46–48.
- Bell W.J., Roth L.M., Nalepa C.A. 2007. Cockroaches: Ecology, Behavior, and Natural
  History. Baltimore: The Johns Hopkins University Press.
- 600 Benton M.J. 2009. The Red Queen and the Court Jester: species diversity and the role of
- 601 biotic and abiotic factors through time. Science 323:728–732.
- Bignell D.E., Eggleton R. 1995. On the elevated intestinal pH of higher termites (Isoptera :
  Termitidae). Insectes Soc. 42:57–69.
- Bourguignon T., Lo N., Cameron S.L., Šobotník J., Hayashi Y., Shigenobu S., Watanabe D.,
- Roisin Y., Miura T., Evans T.A. 2015. The evolutionary history of termites as inferred
  from 66 mitochondrial genomes. Mol. Biol. Evol. 32:406-421.
- 607 Bourke A. 2011. Principles of Social Evolution. Oxford: Oxford University Press.
- Brune A. 2014. Symbiotic digestion of lignocellulose in termite guts. Nat. Rev. Microbiol.
  12:168–180.
- Brune A., Dietrich C. 2015. The gut microbiota of termites: Digesting the diversity in the
  light of ecology and evolution. Annu. Rev. Microbiol. 69:145-166.
- 612 Cardinal S., Danforth B.N. 2011. The antiquity and evolutionary history of social behavior in
  613 bees. PLoS One 6:e21086.
- 614 Cardinal S., Danforth B.N. 2013. Bees diversified in the age of eudicots. Proc. R. Soc. B
  615 280:20122686.

- 616 Chapman R.E., Bourke A.F.G. 2001. The influence of sociality on the conservation biology of
  617 social insects. Ecol. Lett. 4:650–662.
- 618 Cleveland L.R., Hall S.R., Sanders E.P., Collier J. 1934. The wood feeding roach
- *Cryptocercus*, its protozoa, and the symbiosis between protozoa and roach. Mem. Am.
  Acad. Arts Sci. 17:185–342.
- 621 Condamine F.L., Rolland J., Morlon H. 2013. Macroevolutionary perspectives to
  622 environmental change. Ecol. Lett. 16:72–85.
- 623 Condamine F.L., Clapham M.E., Kergoat G.J. 2016. Global patterns of insect diversification:
  624 towards a reconciliation of fossil and molecular evidence? Sci. Rep. 6:19208.
- Darwin C. 1859. On the Origin of Species by Means of Natural Selection, or Preservation of
  Favored Races in the Struggle for Life. London: Murray.
- Davis R.B., Baldauf S.L., Mayhew P.J. 2009. Eusociality and the success of the termites:
  insights from a supertree of dictyopteran families. J. Evol. Biol. 22:1750–1761.
- Davis M.P., Midford P.E., Maddison W. 2013. Exploring power and parameter estimation of
  the BiSSE method for analyzing species diversification. BMC Evol. Biol. 13:38.
- 631 Dupont L., Roy V., Bakkali A., Harry M. 2009. Genetic variability of the soil-feeding termite
- *Labiotermes labralis* (Termitidae, Nasutitermitinae) in the Amazonian primary forest
  and remnant patches. Insect Conserv. Divers. 2:53–61.
- 634 Eggleton P. 2011. An introduction to termites: biology, taxonomy and functional morphology.
- 635 In: Biology of Termites: a Modern Synthesis. (eds. Bignell D.E., Roisin Y., Lo N.).
- 636 Dordrecht: Springer, pp. 1–26.
- 637 Engel M.S., Barden P., Riccio M.L., Grimaldi D.A. 2016. Morphologically specialized
- 638 termite castes and advanced sociality in the Early Cretaceous. Curr. Biol. 26:522-530.

639	Farrell B.D., Sequeira A.S., O'Meara B.C., Normark B.B., Chung J.H., Jordal B.H. 2001. The
640	evolution of agriculture in beetles (Curculionidae: Scolytinae and Platypodinae).
641	Evolution 55:2011-2027.

- FitzJohn R.G., Maddison W.P., Otto S.P. 2009. Estimating trait-dependent speciation and 642
- 643 extinction rates from incompletely resolved phylogenies. Syst. Biol. 58:595-611.
- 644 FitzJohn R.G. 2012. Diversitree : comparative phylogenetic analyses of diversification in R. 645 Methods Ecol. Evol. 3:1084–1092.
- 646 Foster C.S., Sauquet H., Van der Merwe M., McPherson H., Rossetto M., Ho S.Y. 2016.
- 647 Evaluating the impact of genomic data and priors on Bayesian estimates of the 648 angiosperm evolutionary timescale. Syst. Biol. 66:338-351.
- 649 Goodisman M.A.D., Crozier R.H. 2002. Population and colony genetic structure of the 650 primitive termite Mastotermes darwiniensis. Evolution. 56:70-83.
- 651 Goutte S., Dubois A., Howard S.D., Marquez R., Rowley J.J.L., Dehling J.M., Grandcolas P., 652 Xiong R., Legendre F. 2016. Environmental constraints and call evolution in torrent 653 dwelling frogs. Evolution 70:811-826.
- 654 Grandcolas P. 1993. Le genre Paramuzoa Roth, 1973 : sa répartition et un cas de xylophagie 655
- chez les Nyctiborinae (Dictyoptera, Blattaria). Bull. la Soc. Entomol. Fr. 98:131-138.
- 656 Grandcolas P., D'Haese C. 2002. The origin of a "true" worker caste in termites: phylogenetic 657 evidence is not decisive. J. Evol. Biol. 15:885-888.
- 658 Grandcolas P., Bellés X., Piulachs M.D., D'Haese C. 2002. Le genre Lauraesilpha
- 659 Grandcolas 1997 : nouvelles espèces, endémisme, séquences d'ARN ribosomique et
- 660 caractères d'appartenance aux Blattidae (Insectes, Dictyoptères, Blattidae,
- 661 Tyronicinae), in Najt J. & Grandcolas P. (eds), Zoologia Neocaledonica 5. Paris:
- 662 Mémoires du Muséum Natl. d'Histoire Nat. 187:117-131.

- Hall D.W., Goodisman M.A.D. 2012. The effects of kin selection on rates of molecular
  evolution in social insects. Evolution 66:2080–2093.
- Harvey M.G., Seeholzer G.F., Smith B.T., Rabosky D.L., Cuervo A.M., Brumfield R.T. 2017.
- Positive association between population genetic differentiation and speciation rates in
  New World birds. Proc. Natl Acad. Sci. USA 114:6328-6333.
- 668 Heard S.B., Hauser D.L. 1995. Key evolutionary innovations and their ecological
- 669 mechanisms. Hist. Biol. 10:151–173.
- 670 Hines H.M., Hunt J.H., O'Connor T.K., Gillespie J.J., Cameron S.A. 2007. Multigene
- 671 phylogeny reveals eusociality evolved twice in vespid wasps. Proc. Natl Acad. Sci.
- 672 USA 104:3295–3299.
- Inward D., Vogler A.P., Eggleton P. 2007. A comprehensive phylogenetic analysis of termites
  (Isoptera) illuminates key aspects of their evolutionary biology. Mol. Phylogenet.
  Evol. 44:953–967.
- Jablonski D. 2008. Species selection: theory and data. Annu. Rev. Ecol. Evol. Syst. 39:501–
  524.
- 678 Jeanson R., Deneubourg J.-L. 2009. Positive feedback, convergent collective patterns and
- social transitions in arthropods. In: Organization of Insect Societies: From Genome to
  Sociocomplexity (eds Gadau J., Fewell J.). Cambridge, MA, Harvard University Press,
  pp. 460–482.
- Johnstone R.A., Cant M.A., Field J. 2012. Sex-biased dispersal, haplodiploidy and the
  evolution of helping in social insects. Proc. Biol. Sci. 279:787–93.
- Klass K.-D., Nalepa C.A., Lo N. 2008. Wood-feeding cockroaches as models for termite
  evolution (Insecta: Dictyoptera): *Cryptocercus* vs. *Parasphaeria boleiriana*. Mol.
  Phylogenet. Evol. 46:809–817.

687	Köhler T., Dietrich C., Scheffrahn R.H., Brune A. 2012. High-resolution analysis of gut
688	environment and bacterial microbiota reveals functional compartmentation of the gut
689	in wood-feeding higher termites (Nasutitermes spp.). Appl. Environ. Microbiol.
690	78:4691–4701.
691	Korb J. 2009. Termites: an alternative road to eusociality and the importance of group
692	benefits in social insects. In: Organization of Insect Societies: From Genome to
693	Sociocomplexity (eds Gadau J., Fewell J.). Cambridge, MA, Harvard University Press
694	pp. 128–147.
695	Krishna K., Grimaldi D.A., Krishna V., Engel M.S. 2010. Treatise on the Isoptera of the
696	World. Bull. Am. Mus. Nat. Hist. 377:1-2681.
697	Lanfear R., Ho S.Y.W., Love D., Bromham L. 2010. Mutation rate is linked to diversification
698	in birds. Proc. Natl Acad. Sci. USA 107:20423-20428.
699	Lanfear R., Kokko H., Eyre-Walker A. 2014. Population size and the rate of evolution.
700	Trends Ecol. Evol. 29:33–41.
701	LaPolla J.S., Dlussky G.M., Perrichot V. 2013. Ants and the fossil record. Annu. Rev.
702	Entomol. 58:609-630.
703	Legendre F., Deleporte P., Depraetere M., Gasc A., Pellens R., Grandcolas P. 2015a. Dyadic
704	behavioural interactions in cockroaches (Blaberidae): ecomorphological and
705	evolutionary implications. Behaviour 152:1229-1256.
706	Legendre F., Nel A., Svenson G.J., Robillard T., Pellens R., Grandcolas P. 2015b. Phylogeny

- of Dictyoptera: Dating the origin of cockroaches, praying mantises and termites with
  molecular data and controlled fossil evidence. PLoS One 10:e0130127.
- 709 Legendre F., Whiting M.F., Bordereau C., Cancello E.M., Evans T.A., Grandcolas P. 2008.
- 710 The phylogeny of termites (Dictyoptera: Isoptera) based on mitochondrial and nuclear

- 711 markers: Implications for the evolution of the worker and pseudergate castes, and
  712 foraging behaviors. Mol. Phylogenet. Evol. 48:615–627.
- 713 Legendre F., Whiting M.F., Grandcolas P. 2013. Phylogenetic analyses of termite post-
- 714 embryonic sequences illuminate caste and developmental pathway evolution. Evol.
  715 Dev. 15:146–157.
- 716 Lepage M., Darlington J.P.E.C. 2000. Population dynamics of termites. In: Termites:
- 717 Evolution, Sociality, Symbioses, Ecology (eds. Abe, T. Bignel, D.E., Higashi, M.).
  718 Dordrecht: Springer, pp. 333–361.
- T19 Liow L.H., Reitan T., Harnik P.G. 2015. Ecological interactions on macroevolutionary time

scales: clams and brachiopods are more than ships that pass in the night. Ecol. Lett.
18:1030–1039.

- Maddison W.P., Midford P.E., Otto S.P. 2007. Estimating a binary character's effect on
  speciation and extinction. Syst. Biol. 56:701–710.
- 724 Magallón S., Gómez-Acevedo S., Sánchez-Reyes L.L., Hernández-Hernández T. 2015. A
- metacalibrated time-tree documents the early rise of flowering plant phylogeneticdiversity. New Phytol. 207:437-453.
- Mikaelyan A., Dietrich C., Köhler T., Poulsen M., Sillam-Dussès D., Brune A. 2015. Diet is
  the primary determinant of bacterial community structure in the guts of higher
  termites. Mol. Ecol. 24:5284–5295.
- 730 Moore B.R., Höhna S., May M.R., Rannala B., Huelsenbeck J.P. 2016. Critically evaluating
- the theory and performance of Bayesian analysis of macroevolutionary mixtures. Proc.
  Natl Acad. Sci. USA 113:9569–9574.
- 733 Moreau C.S., Bell C.D., Vila R., Archibald S.B., Pierce N.E. 2006. Phylogeny of the ants:
- diversification in the age of angiosperms. Science 312:101-104.

- Morlon H., Lewitus E., Condamine F.L., Manceau M., Clavel J., Drury J. 2016. RPANDA:
  An R package for macroevolutionary analyses on phylogenetic trees. Methods Ecol.
  Evol. 7:589–597.
- Morlon H., Parsons T.L., Plotkin J.B. 2011. Reconciling molecular phylogenies with the
  fossil record. Proc. Natl Acad. Sci. USA 108:16327-16332.
- 740 Morlon H., Potts M.D., Plotkin J.B. 2010. Inferring the dynamics of diversification: a

coalescent approach. PLoS Biol. 8:e1000493.

- Nee S. 2006. Birth-death models in macroevolution. Annu. Rev. Ecol. Evol. Syst. 37:1-17.
- Noirot C., Pasteels J.M. 1987. Ontogenetic development and evolution of the worker caste in
  termites. Experientia 43:851–952.
- Nonacs P. 2011. Kinship, greenbeards, and runaway social selection in the evolution of social
  insect cooperation. Proc. Natl Acad. Sci. USA 108:10808–10815.
- 747 Nowak M.A., Tarnita C.E., Wilson E.O. 2010. The evolution of eusociality. Nature
  748 466:1057–1062.
- 749 Okasha S. 2006. The levels of selection debate: philosophical issues. Philos. Compass 1:74–
  750 85.
- 751 Pellens R., D'Haese C.A., Bellés X., Piulachs M.D., Legendre F., Wheeler W.C., Grandcolas
- P. 2007. The evolutionary transition from subsocial to eusocial behaviour in
- Dictyoptera: Phylogenetic evidence for modification of the "shift-in-dependent-care"
  hypothesis with a new subsocial cockroach. Mol. Phylogenet. Evol. 43:616–626.
- Pellens R., Grandcolas P., da Silva-Neto I.D. 2002. A new and independently evolved case of
- xylophagy and the presence of intestinal flagellates in the cockroach *Parasphaeria*
- 757 *boleiriana* (Dictyoptera, Blaberidae, Zetoborinae) from the remnants of the Brazilian
- 758 Atlantic forest. Can. J. Zool. 80:350–359.

759	Peters R.S., Krogmann L., Mayer C., Donath A., Gunkel S., Meusemann K., Kozlov A.,
760	Podsiadlowski L., Petersen M., Lanfear R., Diez P.A., Heraty J., Kjer K.M.,
761	Klopfstein S., Meier R., Polidori C., Schmitt T., Liu S., Zhou X., Wappler T., Rust J.,
762	Misof B., Niehuis O. 2017. Evolutionary history of the Hymenoptera. Curr. Biol.
763	27:1013-1018.
764	Phillimore A.B., Price T.D. 2008. Density-dependent cladogenesis in birds. PLoS Biol. 6:e71.
765	Pie M.R., Feitosa R.M. 2016. Relictual ant lineages (Hymenoptera: Formicidae) and their
766	evolutionary implications. Myrmecological News 22:55-58.
767	Rabosky D.L. 2009. Ecological limits and diversification rate: alternative paradigms to
768	explain the variation in species richness among clades and regions. Ecol. Lett. 12:735-
769	743.
770	Rabosky D.L. 2014. Automatic detection of key innovations, rate shifts, and diversity-
771	dependence on phylogenetic trees. PLoS One 9:e89543.
772	Rabosky D.L., McCune A.R. 2010. Reinventing species selection with molecular
773	phylogenies. Trends Ecol. Evol. 25:68–74.
774	Rabosky D.L., Matute D.R. 2013. Macroevolutionary speciation rates are decoupled from the
775	evolution of intrinsic reproductive isolation in Drosophila and birds. Proc. Natl Acad.
776	Sci. USA 110:15354–15359.
777	Rabosky D.L., Goldberg E.E. 2015. Model inadequacy and mistaken inferences of trait-
778	dependent speciation. Syst. Biol. 64:340-355.
779	Rabosky D.L., Santini F., Eastman J., Smith S.A., Sidlauskas B., Chang J., Alfaro M.E. 2013.
780	Rates of speciation and morphological evolution are correlated across the largest
781	vertebrate radiation. Nat. Commun. 4:1958.

- 782 Rabosky D.L., Grundler M., Anderson C., Title P., Shi J.J., Brown J.W., Huang H., Larson
- J.G. 2014. BAMMtools: An R package for the analysis of evolutionary dynamics on
  phylogenetic trees. Methods Ecol. Evol. 5:701–707.
- Rabosky D.L., Mitchell J.S., Chang J. 2017. Is BAMM flawed? Theoretical and practical
  concerns in the analysis of multi-rate diversification models. Syst. Biol. 66:477-498.
- 787 Rainford J.L., Mayhew P.J. 2015. Diet evolution and clade richness in Hexapoda: a

phylogenetic study of higher taxa. Am. Nat. 186:777-791.

- 789 Rainford J.L., Hofreiter M., Nicholson D.B., Mayhew P.J. 2014. Phylogenetic distribution of
- extant richness suggests metamorphosis is a key innovation driving diversification ininsects. PLoS One 9:e109085.
- Raymond A., Cutlip P., Sweet M. 2000. Rates and processes of terrestrial nutrient cycling in
- the Paleozoic: the world before beetles, termites, and flies. In: Evolutionary
- Paleoecology: the Ecological Context of Macroevolutionary Change (eds Allmon W.,

Bottjer D.). New York: Columbia University Press, pp. 235–283.

- Rehan S.M., Toth A.L. 2015. Climbing the social ladder: the molecular evolution of sociality.
  Trends Ecol. Evol. 30:426–433.
- Roisin Y., Korb J. 2011. Social Organisation and the Status of Workers in Termites. In:
- Biology of Termites: a Modern Synthesis (eds Bignell D.E., Roisin Y., Lo N.).
- 800 Dordrecht: Springer, pp. 133–164.
- Rousset F., Lion S. 2011. Much ado about nothing: Nowak et al.'s charge against inclusive
  fitness theory. J. Evol. Biol. 24:1386–1392.
- 803 Sánchez-García M., Matheny P.B. 2016. Is the switch to an ectomycorrhizal state an
- 804 evolutionary key innovation in mushroom-forming fungi? A case study in the
- 805 Tricholomatineae (Agaricales). Evolution 71:51-65.

806	Silvestro D., Antonelli A., Salamin N., Quental T.B. 2015a. The role of clade competition in
807	the diversification of North American canids. Proc. Natl Acad. Sci. USA 112:8684-
808	8689.

809 Silvestro D., Cascales-Miñana B., Bacon C.D., Antonelli A. 2015b. Revisiting the origin and

- 810 diversification of vascular plants through a comprehensive Bayesian analysis of the
  811 fossil record. New Phytol. 207:425-436.
- 812 Simpson G.G. 1944. Tempo and Mode in Evolution. New York: Columbia University.
- 813 Thompson G.J., Kitade O., Lo N., Crozier R.H. 2000. Phylogenetic evidence for a single,

ancestral origin of a "true" worker caste in termites. J. Evol. Biol. 13:869–881.

815 Thompson G.J., Lenz M., Crozier R.H., Crespi B.J. 2007. Molecular-genetic analyses of

816 dispersal and breeding behaviour in the Australian termite *Coptotermes lacteus*:

- 817 Evidence for non-random mating in a swarm-dispersal mating system. Aust. J. Zool.
  818 55:219–227.
- 819 Vargo E.L., Husseneder C. 2011. Genetic structure of termite colonies and populations. In:

Biology of Termites: a Modern Synthesis (eds Bignell D.E., Roisin Y., Lo N.).

- B21 Dordrecht: Springer, pp. 321–347.
- 822 Ware J.L., Grimaldi D.A., Engel M.S. 2010. The effects of fossil placement and calibration

823 on divergence times and rates: an example from the termites (Insecta: Isoptera).

824 Arthropod Struct. Develop. 39:204-219.

825 Watson J.A.L., Sewell J.J. 1985. Caste development in *Mastotermes* and *Kalotermes*: which

- 826 is primitive? In: Caste Differentiation in Social Insects (eds Watson J.A.L., Okot-
- 827 Kotber B.M., Noirot C.). Book Section. Pergamon, Oxford, pp. 27–40.
- Webster A.J., Payne R.J.H., Pagel M. 2003. Molecular phylogenies link rates of evolution and
  speciation. Science 301:478–478.

- Wilson E.O. 1971. The Insect Societies. The Belknap Press of the Harvard University Press,
  Cambridge, Massachusetts, 548 p.
- 832 Wilson E.O. 1990. Success and Dominance in Ecosystems: the Case of the Social Insect.

Ecology Institute, Oldendorf-L, Germany, 104 p.

- Wilson E.O. 1992. The effects of complex social life on evolution and biodiversity. Oikos63:13–18.
- Wilson E.O., Hölldobler B. 2005. Eusociality: origin and consequences. Proc. Natl Acad. Sci
  USA 102:13367–13371.

Models NP logL AICc ΔAIC AICω Subclade speciation rate Subclade extinction rate Subclade net div. rate		Scenario with a rate shift located at:											
Models	Whole phylogeny	the MRCA of Isoptera	the MRCA of Euisoptera	the MRCA of Kalotermitidae and Neoisoptera	the MRCA of Neoisoptera	the MRCA of Neoisoptera minus Rhinotermitinae	the MRCA of Termitidae, Coptotermitinae and Heterotermitinae	the MRCA of Termitidae					
NP	3	4	4	5	4	5	5	4					
logL	-3889.247	-3709.158	-3706.857	-3693.107	-3658.84	-3659.554	-3651.496	-3691.284					
AICc	7784.526	7426.385	7421.781	7396.308	7325.754	7329.206	7313.092	7390.653					
ΔΑΙC	471.434	113.293	108.689	83.216	12.662	16.114	0	77.561					
AICω	0	0	0	0	≈0.002	< 0.0001	0.998	0					
Subclade speciation rate	0.191	0.177	0.178	0.185	0.186	0.191	0.193	0.197					
Subclade extinction rate	0.175	0	0	0	0	0	0	0					
Subclade net div. rate	0.016	0.177	0.178	0.185	0.186	0.191	0.193	0.197					
Backbone speciation rate	-	0.044	0.044	0.057	0.044	0.058	0.058	0.084					
Backbone extinction rate	-	0	0	0.024	0	0.024	0.023	0.06					
Backbone net. div. rate	-	0.044	0.044	0.033	0.044	0.034	0.035	0.024					

TABLE 1.	Support for a	n increase c	of diversification	on rates for	termites.	especially	for lineas	ges including	'true'	workers.
	~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~									

*Notes:* The table reports the log-likelihood (*logL*) and the corrected Akaike Information Criterion (*AICc*) calculated for the whole phylogeny of Dictyoptera (assuming no shift of diversification) and seven evolutionary scenarios including a shift of diversification (as specified in the table). For each model, the parameter values of the best-fitting model are reported (rates are in events/Myr). The 'backbone' rates represent the estimated diversification parameters for the tree containing all Dictyoptera species except the termite subclades. The termite subclades are analyzed aside. The *logL*, the number of parameters (*NP*), and *AICc* are thus the sum of the logL/parameters/AICc of the backbone tree plus the logL/parameters/AICc of the termite subclade. The best-fitting scenario is determined by  $\Delta AIC$  and  $AIC \omega$  and includes a shift within the termites that only include species with 'true' workers in their societies (*scenario 7*, highlighted in bold).

TABLE 2. Supports for eusociality as a driver of diversification

Models	NP	logL	AICc	ΔAICc	AICω	λ0	λ1	μ0	μ1	q01	q10		
Null model	3	-3831.072	7668.175	218.2023	0	0.1980		0.1786		0.1980 0.1786		2.01	E-05
λ0≠λ1, μ0=μ1, q01=q10	4	-3752.206	7512.465	62.492	0	0.1335 0.1867		0.1129		1335 0.1867 0.1129 1.0		1.64	E-05
λ0=λ1, μ0≠μ1, q01=q10	4	-3773.267	7554.586	104.6133	0	0.1626		0.1441	0.0921	1.24	E-05		
λ0=λ1, μ0=μ1, q01≠q10	4	-3830.967	7669.988	220.0149	0	0.19	0.1979		0.1784		4.97E-06		
λ0≠λ1, μ0≠μ1, q01=q10	5	-3719.947	7449.973	0	0.887	0.1004	0.4145	0.0766	0.3721	0.3	721		
λ0≠λ1, μ0=μ1, q01≠q10	5	-3752.346	7514.772	64.7993	0	0.1280	0.1815	0.10	)71	1.81E-05	1.20E-05		
λ0=λ1, μ0≠μ1, q01≠q10	5	-3774.607	7559.294	109.3214	0	0.1541		0.1349	0.0810	1.27E-05	7.40E-06		
λ0≠λ1, μ0≠μ1, q01≠q10	6	-3720.997	7454.106	4.1336	0.113	0.0910	0.4099	0.0667	0.3674	4.35E-05	5.91E-06		

*Notes:* The table reports the models applied, their number of parameters (*NP*), the means for the log-likelihood (*logL*) and the corrected Akaike Information Criterion (*AICc*), the difference of AICc between the best model (lowest AIC) and a given model ( $\Delta AICc$ ), the Akaike weight (*AIC* $\omega$ ) and the values for each parameter of the corresponding model based on 100 dated trees. The best model is highlighted in bold. Standard errors for each parameter estimate are reported in Table S2.  $\theta$  = non-eusocial and I = eusocial;  $\lambda$  = speciation rate (one for each character state,  $\lambda I$  is the speciation rate for eusocial species);  $\mu$  = extinction rate (one for each character state,  $\mu I$  is the extinction rate for eusocial species); and q = transition rate between character states.

TABLE 3. Supports for eusociality with true workers as drivers of diversification

Models	NP	logL	AICc	ΔΑΙΟ	ΑΙϹω	λ1	λ2	λ3	μ1	μ2	μ3	q12	q13	q21	q23	q31	q32
Null model	3	-3889.534	7785.100	373.94	0		0.1719			0.1525				4.2	1E-05		
all $\lambda$ are varying	5	-3717.017	7444.114	32.95	0	0.0683	0.0727	0.1699		0.0438				2.4	3E-05		
all $\mu$ are varying	5	-3751.789	7513.657	102.49	0		0.1214		0.1020	0.0974	0.0000		1.43E-05				
all q are varying	8	-3879.999	7776.191	365.03	0		0.1713			0.1519		1.29E- 05	6.67E- 07	1.67E- 06	0.00161	4.64E- 06	0.00023
all $\lambda$ and $\mu$ are varyin	g 7	-3706.089	7426.327	15.16	≈0.005	0.0637	0.0580	0.3198	0.0384	0.0266	0.2267		3.85E-05				
all $\lambda$ and q are varyin	g 10	-3703.355	7427.005	15.84	≈0.005	0.0645	0.0693	0.1736		0.0394		1.55E- 05	1.33E- 07	7.90E- 07	0.00155	9.81E- 06	2.73E- 06
all $\mu$ and q are varyin	g 10	-3738.133	7496.560	85.40	0		0.1258		0.1067	0.1030	0.0000	1.87E- 05	7.14E- 08	2.63E- 07	0.00043	2.73E- 06	0.00044
All rates are free	12	-3693.372	7411.163	0	0.99	0.0649	0.0378	0.3196	0.0398	0.0000	0.2244	1.09E- 05	2.43E- 07	6.45E- 07	0.00224	1.32E- 06	0.00027

*Notes:* I = non-eusocial or societies without worker, 2 = societies with pseudergates, and 3 = societies with true workers;  $\lambda =$  speciation rate (one for each character state,  $\lambda I$  is the speciation rate for species without worker);  $\mu =$  extinction rate (one for each character state,  $\mu I$  is the extinction rate for species without worker); and q = transition rate between character states.

Models	NP	logL	AICc	ΔΑΙC	AICω	λ1	λ2	λ3	μ1	μ2	μ3	q12	q13	q21	q23	q31	q32
Null model	3	-3840.575	7687.181	274.01	0		0.1959			0.1764				3.03	E-05		
all $\lambda$ are varying	5	-3710.247	7430.573	17.41	0	0.1166	0.1270	0.2157		0.0941				1.72	E-05		
all $\mu$ are varying	5	-3731.518	7473.115	59.95	0		0.1642		0.1456	0.1373	0.0447		1.21E-05				
all q are varying	8	-3835.418	7687.028	273.86	0		0.1957			0.1763		2.40E-05	6.75E-07	4.36E-06	0.00046	1.88E-06	3.10E- 06
all λ and μ are varying	7	-3699.510	7413.168	0	0.703	0.1070	0.1894	0.3022	0.0832	0.1652	0.1979			2.39	E-05		
ll $\lambda$ and q are varying	10	-3703.777	7427.847	14.68	0	0.1147	0.1254	0.2157		0.0921		1.98E-05	2.74E-07	3.45E-06	0.00025	3.21E-06	2.81E- 06
ll $\mu$ and q are varying	10	-3735.700	7491.694	78.53	0		0.1683		0.1500	0.1304	0.0555	2.01E-05	7.33E-07	3.67E-06	0.01504	2.54E-06	0.0022
All rates are free	12	-3695.237	7414.892	1.72	0.297	0.1037	0.1730	0.3033	0.0795	0.1472	0.1975	3.11E-05	4.02E-07	4.87E-06	0.00029	4.52E-06	2.54E- 06

TABLE 4. Supports for gut microbiota as drivers of diversification

*Notes:* I = gut with no specialized microbiota for lignocellulose digestion,  $2^* = \text{gut}$  with cellulolytic flagellates, and  $3^* = \text{gut}$  with entirely prokaryotic microbiota;  $\lambda = \text{speciation}$  rate (one for each character state,  $\lambda I$  is the speciation rate for species without microbiota);  $\mu = \text{extinction}$  rate (one for each character state,  $\mu I$  is the extinction rate for species without microbiota); and q = transition rate between character states. \* Results of alternative coding strategies (see main text) are provided in Figs. S11-S12.





23 2.6 Million years ago (Ma)

a) Evolutionary history of Dictyoptera

